

Article



Revision of Palaearctic and Oriental *Necrophila* Kirby & Spence, part 1: subgenus *Deutosilpha* Portevin (Coleoptera: Silphidae)

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Abstract

Taxonomic revision of the subgenus *Deutosilpha* Portevin, 1920 (of *Necrophila* Kirby & Spence, 1828) through southeastern Asia is presented. Two species are recognised: (1) *N. (D.) rufithorax* (Wiedemann, 1823), comb. nov. (ex *Deutosilpha*), with *Oiceoptoma tetraspilotum* Hope, 1833 as confirmed junior subjective synonym, from India, Nepal and Sri Lanka; and (2) *N. (D.) luciae* Růžička & Schneider, **sp. nov.** from Thailand, Laos, Vietnam and China: Sichuan province. A lectotype is designated for *Silpha rufithorax* Wiedemann, 1823 and *Oiceoptoma tetraspilotum* Hope, 1833. Morphology-based diagnosis and key to adults of both species are produced. Georeferenced records for both species are mapped.

Key words: Coleoptera, Silphidae, *Necrophila*, *Deutosilpha*, taxonomy, new species, new combination, distribution, Oriental region

Introduction

Portevin (1920) erected *Deutosilpha* as a subgenus of *Eusilpha* Semenov, 1890, to accommodate only one species, originally described as *Silpha rufithorax* by Wiedemann (1823). Later, Portevin (1926) treated *Deutosilpha* as a separate genus, related to Oriental and Eastern Palaearctic *Calosilpha* Portevin, 1920, *Chrysosilpha* Portevin, 1921 and *Eusilpha*, and redescribed all four genera. Hatch (1928) catalogued *Deutosilpha* as a subgenus of widely interpreted *Silpha* Linnaeus, 1758. Only recently, Peck (2001) and Sikes (2008) listed *Deutosilpha* along with *Calosilpha*, *Chrysosilpha* and *Eusilpha* as subgenera of Nearctic *Necrophila* Kirby & Spence, 1828; following unpublished taxonomical revision of A.F. Newton, Jr. This classification is also accepted here, detailed phylogenetic relationships of all subgenera will be treated in a separate study (J. Růžička, unpublished).

Phylogenetic affinities of these five genera/subgenera are not clear. Recently, Ikeda *et al.* (2008) published phylogenetic reconstruction of Silphinae based on sequences of one mitochondrial (16S) and three nuclear genes (28S, wingless (Wg), and phosphoenolpyruvate carboxykinase (PepCK)). Both Bayesian and maximum parsimony analyses produced the following tree topology: *Necrophila* + (*Eusilpha* + (*Calosilpha* + *Chrysosilpha*)) (Ikeda *et al.* 2008: 2072, fig. 1). However, *Deutosilpha* was not included into this analysis.

In another paper, Ikeda *et al.* (2007) studied flight capabilities and feeding habits of several species of Japanese carrion beetles of the subfamily Silphidae. *Necrophila* (*Calosilpha*) *brunnicollis* (Kraatz, 1877) and *N.* (*Eusilpha*) *jakowlewi* (Semenov, 1890) have fully developed flight muscles, but *N.* (*E.*) *japonica* (Motschulsky, 1861) showed flight muscle dimorphism. Comparing carbon and nitrogen stable isotope ratios based on the entire beetle body with other beetle predators and necrophages (Coleoptera: Carabidae and Silphidae), they concluded that *N.* (*C.*) *brunnicollis* is necrophagous, but *N.* (*E.*) *jakowlewi* and *N.* (*E.*) *japonica* are predaceous (interpreted as derived state from necrophagy; Ikeda et al. 2008: 2075, fig. 3).

The aim of this paper is to redescribe the subgenus *Necrophila* (*Deutosilpha*), Oriental in distribution, and to name a new species, separate from the type species of *Deutosilpha*, based on study of material from the eastern part of the distributional range of the subgenus. Detailed distribution of both species is also summarized and mapped, based on material available from museums and private collection.

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Material and methods

Specimens examined in this study are deposited in the following museums and private collections (acronyms according to Arnett *et al.* 1993):

BMNH Natural History Museum, London (R. Booth, M.V.L. Barclay); **FMNH** Field Museum of Natural History, Chicago (A.F. Newton, J.H. Boone); **HNHM** Magyar Természettudományi Muzeum, Budapest (O. Merkl); **JHAC** collection of Jiří Háva, Únětice u Prahy, Czech Republic; **JMIC** collection of Jiří Mička, Praha, Czech Republic; **JRUC** collection of Jan Růžička, Praha, Czech Republic; **JSCC** collection of Jan Schneider, Praha, Czech Republic; Muséum d'Histoire Naturelle, Genève (G. Cuccodoro); MHNG **MNHN** Muséum national d'Histoire naturelle, Paris (†Nicole Berti, Azadeh Taghavian); **MNIC** collection of M. Nishikawa, Ebina, Japan; **MTDC** collection of Miloš Trýzna, Děčín, Czech Republic; Naturhistorisches Museum, Basel (M. Brancucci, Eva Sprecher-Uebersax); NHMB Naturhistorisches Museum, Wien (H. Schönmann); **NHMW NHRS** Naturhistoriska riksmuseet, Stockholm (B. Viklund); Národní museum, Praha (J. Jelínek, J. Hájek); **NMPC OUMNH** Oxford University Museum of Natural History, Oxford (D.J. Mann); **RMNH** National Museum of Natural History Naturalis, Leiden (A. von Assen); **SMFD** Forschungsinstitut Senckenberg, Frankfurt am Main (Andrea Vesmanis, D. Kovac); **SMLC** Severočeské muzeum, Liberec, Czech Republic (P. Vonička); Staatliches Museum für Naturkunde, Stuttgart (W. Schawaller); **SMNS TFUC** collection of T. Fukuzawa, Kanagawa, Japan; collection of W. Barries, Wien, Austria; **WBAC ZMAN** Zoölogisch Museum Amsterdam, Amsterdam (S.A. Ulenberg, W. Hogenes); **ZMAS** Russian Academy of Sciences, Zoological Institute, St. Petersburg (M.G. Volkovich); Zoological Museum, University of Copenhagen, Copenhagen (A.Yu. Solodovnikov); **ZMUC ZSM** Zoologische Staatssammlung, München (M. Balke).

Exact label data are cited only for the type material. Separate lines on labels are indicated (only for primary types) by "/", separate labels by "/". Authors' remarks and comments are enclosed in square brackets. [p] – preceding data are printed; [h] – preceding data are hand-written.

Photographs were taken using an Olympus SZX12 stereomicroscope with Olympus E-330 camera attached and multiple (5–23) layers of focus combined in the Helicon Focus 4.21 software. Male and female terminalia were studied after short clearing in hot KOH and mounted in temporal glycerine mounts for line drawings. Terminology of male genital segment follows mostly Blackburn (1936).

Microsoft Encarta Premium 2008 (Microsoft Corporation 2007), NGA GEOnet Names Server (National Geospatial-intelligence Agency 2009; http://earth-info.nga.mil/gns/html/index.html), Google Earth (Google 2010; http://earth.google.com) and Fuzzy Gazetteer (Ch. Kohlschütter 2009; http://isodp.fh-hof.de/fuzzyg/query/) were used to find coordinates for most of the localities. Distribution map was produced using Online Map Creation (Weinelt 2006; www.aquarius.ifm-geomar.de).

Results

subgenus Necrophila (Deutosilpha) Portevin, 1920

Deutosilpha Portevin, 1920: 396 (description, as subgenus of Eusilpha, type species: Silpha rufithorax Wiedemann, 1823, by monotypy)

Deutosilpha: Portevin, 1926: 108, 151 (redescription, as separate genus, catalogue)

Silpha (Deuterosilpha): Hatch 1928: 112 (catalogue, subsequent misspelling of Deutosilpha, as subgenus of Silpha)

Necrophila (Deutosilpha): Peck 2001: 270 (catalogue, as subgenus of Necrophila) Necrophila (Deutosilpha): Sikes 2008: 752 (catalogue, as subgenus of Necrophila)

Diagnostic description. Body flattened, length 17.5–22.5 mm. Head black with blue metallic lustre and yellow gula, antennae and mouthparts black. Pronotum orange with four black, glabrous spots, arranged in semi-circular position posteriorly on disc (Figs. 13–16). Scutellum, elytra, meso- and metathorax and legs black with blue metallic lustre. Abdomen black with metalic lustre, with pale posterior part of ventrite 8 and pale segments 9 and 10. Elytra dark blue to violet on outer lateral margins (on elytral epipleura), and light metallic blue on ventral surface (which is exposed probably only during flight).

Head flattened, with row of long erect black setae behind the eyes (Fig. 22). From with weakly indicated fovea between the eyes. Eyes small, not prominently protruding (Figs. 21–22). Antennae with loosely formed antennal club, consisting of 4 antennomeres (Fig. 15).

Pronotum hexagonal in shape (Figs. 21–22), with anterior margin only weakly emarginte, posterior margin straight medially (Figs. 21–22). Surface with uniformly dispersed, fine and dense, round punctures; more superficial on pronotal disc, without setation. Surface matt, with very fine isodiametrical microsculpture. Anterior and posterior margin with dense row of stout, short, yellow, ventrally oriented setae.

Scutellum large, with sinuous lateral margins (Figs. 13–16), densely covered by fine round punctures.

Elytra flattened with dorsally elevated, robust epipleural ridges on lateral margins. Each elytron with two weakly developed costae and abbreviated, outer third costa, which is only vestigial (Figs. 13–16). Elytral costae more distinctly delimited and elevated in females (Figs. 14, 16) than in males (Figs. 13, 15). Elytral epipleural ridge distinctly elevated along lateral margin, forming crest-like structures abruptly flattened postero-laterally in males (Figs. 13, 15; 17, 19), and gradually diminishing in females (Figs. 14, 16; 18, 20). Apex of elytron in males subtruncate to truncate with sinuous margin (Figs. 17, 19), in females elongate to slender, triangular or sinuate tip, sometimes with suture extended in distinct inner denticle (Figs. 18, 20). Dorsal surface with uniformly dispersed, fine and dense punctation, similar to that on pronotum; surface matt, with very fine, isodiametric microsculpture; without reticulate sculpture (which is present in *Necrophila* s. str.). Ventrally, elytra covered by densely arranged, large and strong punctures.

Hind wings fully developed, functional.

Metaventrite with dense, black pubescence and fine, transverse microsculpture.

Legs with bent tibia (more pronounced in males), tibia armed with several longitudinal rows of short, strong setae. Pro- and mesofemur with distal, ventrally oriented denticle in males (unmodified in females). Protarsomere 1–4 moderately expanded in males (unmodified in females). Metatarsal claws asymmetrical in males (Fig. 27), with inner claw distinctly longer and more robust than the outer one (almost symmetrical, of the comparable size in females, Fig. 28).

Abdomen very tightly fixed by ventrally expanded elytral epipleura.

Male. Genital segment with longitudinal ventrite 9, reaching anterior margin of tergum 9 (Figs. 3, 8); spiculum gastrale medium in size, relatively slender, slightly asymmetrical (Figs. 2–3, 7–8). Aedeagus small; with slender paramerae and median lobe regularly tapering to a triangular tip (Figs. 1, 6).

Female. Genital segment with tergum 10 hexagonal in shape in dorsal view (Figs. 4, 9, 11), with stylus inserted laterally, shorter than apex of coxite in ventral view (Figs. 5, 10, 12).

A key to the species of Necrophila (Deutosilpha)

- Body more slender (Figs. 13–14), with pronotum 1.35–1.57 times as wide as long (Fig. 21). Apex of elytron in male subtruncate, with regularly rounded posterior margin (Fig. 17); in female elongate, with triangular, flat apex, only with vestigial inner denticle or without denticle (Fig. 18). (Thailand, Laos, Vietnam, China: Sichuan province; Fig. 33).... N. (D.) luciae sp. nov.

Necrophila (Deutosilpha) rufithorax (Wiedemann, 1823), comb. nov.

(Figs. 1–5, 15–16, 19–20, 22–28, 33)

Silpha rufithorax Wiedemann, 1823: 71 (description, type locality "Bengalia")

Silpha rufithorax: Harold 1877: 347 (senior subjective synonym of Silpha tetraspilota)

Eusilpha rufithorax: Portevin 1903: 332 (new combination, confirmed as senior subjective synonym of Silpha tetraspilota)

Eusilpha rufithorax: Portevin 1905: 51 (synonymy confirmed)

Silpha rufithorax: Arrow 1909: 190 (new combination, synonymy confirmed)

Eusilpha (Deutosilpha) rufithorax: Portevin 1920: 396 (new subgeneric assignment)

Deutosilpha rufithorax: Portevin 1926: 111, 151 (new combination, redescription, catalogue)

Silpha (Deuterosilpha) rufithorax: Hatch 1928: 112 (catalogue)

Oiceoptoma tetraspilotum Hope, 1833: 61 (description, type locality "Indiâ Orientali circa Poona")

Oiceoptoma tetraspilotum Hope, 1834: 93 (redescription)

Silpha tetraspilota: Harold 1877: 347 (new combination, junior subjective synonym of Silpha rufithorax)

Eusilpha tetraspilota: Portevin 1903: 332 (new combination, synonymy confirmed)

Eusilpha tetraspilota: Portevin 1905: 51 (synonymy confirmed)

Silpha tetraspilota: Arrow 1909: 190 (new combination, synonymy confirmed)

Deutosilpha tetraspilota: Portevin 1926: 151 (new combination, synonymy confirmed, catalogue)

Silpha (Deuterosilpha) tetraspilota: Hatch 1928: 112 (new combination, catalogue)

Type material examined. Lectotype ♂ of Silpha rufithorax (ZMUC) (here designated), labelled (Fig. 23): "Mus. / Westerm. [p] // TYPE [p, red label] // S. rufithorax / Wied. / Bengal / May 1809 [hw] // LECTOTYPUS / Silpha rufithorax / Wiedemann, 1823 / Jan Růžička & / Jan Schneider des. 2009 [p, red label] // Necrophila (Deutosilpha) / rufithorax / (Wiedemann, 1823) [p] ♂ [hw] / Jan Růžička det. 200 [p] 9 [hw]"; paralectotype ♀ (ZMUC), labelled: "Mus. / Westerm. [p] // TYPE [p, red label] // PARALECTOTYPUS / Silpha rufithorax / Wiedemann, 1823 / Jan Růžička & / Jan Schneider des. 2009 [p, red label] // Necrophila (Deutosilpha) / rufithorax / (Wiedemann, 1823) / Jan Růžička det. 200 [p] 9 [hw]".

Lectotype ♀ of *Oiceoptoma tetraspilotum* (BMNH) (here designated), labelled (Fig. 24): "Type [p, red rim, round label] // Duk= / hun [hw, round label] [= Deccan plateau] / Col. Sykes [hw, on underside] // Silpha / tetraspilota / Type. Hope. [hw] // Ind. Mus. / 79.64. [p] // LECTOTYPUS / Oiceoptoma / tetraspilotum Hope, 1833 / Jan Růžička & / Jan Schneider des. 2010 [p, red label] // Necrophila (Deutosilpha) / rufithorax / (Wiedemann, 1823) [p] ♀ [hw] / Jan Růžička det. 200 [p] 9 [hw]"; Paralectotypes: ♂ (OUMNH), labelled (Fig. 25): "TYPE / HOPE [p] / Trans. Zool. / Lond. 1. P.93 / T. 13 fig. 3 [hw] / Coll. Hope Oxon. [p, red frame] // Oiceoptoma / 4spilotum / Hope J. Z. L [Hope's hw, red label] // TYPE [p] COL : 237 1/2 / Oiceoptoma [sic!] / tetraspilota Hope [hw] / HOPE DEPT. OXFORD [p, black frame] // PARALECTOTYPUS / Oiceoptoma / tetraspilotum Hope, 1833 / Jan Růžička & / Jan Růžička det. 201 [p] 0 [hw]"; 1 ♀ (OUMNH), labelled (Fig. 26): "tetraspilotum / Hope [Hope's hw] // TYPE [p] COL : 237 2/2 / Oiceoptoma [sic!] / tetraspilota Hope [hw] / HOPE DEPT. OXFORD [p, black frame] // PARALECTOTYPUS / Oiceoptoma / tetraspilotum Hope, 1833 / Jan Růžička & / Jan Schneider des. 2010 [p, red label] // Necrophila (Deutosilpha) / rufithorax / (Wiedemann, 1823) [p] ♀ [hw] / Jan Růžička det. 201 [p] 0 [hw]".

 and collector's name, $1 \subsetneq (ZMAN)$; same locality, without date and collector's name, $1 \circlearrowleft 1 \subsetneq (RMNH)$; same locality, "13748", without date [but before 1905], ex coll. Fry, 1 \(\text{(BMNH)} \); same locality, without date, ex coll. Mniszech, $1 \, \circlearrowleft$, $2 \, \subsetneq \varphi$ (coll. A. Grouvelle, MNHN); Poona [= Pune, ca. 18°31'N 073°50'E], 60.15 E.I.C., without date and collector's name [but before 1860], 1 ♂, 1 ♀ (BMNH); same locality, 26.viii.1944, D. Leston leg., 2 ♂♂, 1 \subsetneq (BMNH); Pune, without date [but before 1917] and collector's name, 1 \subsetneq (coll. A. Grouvelle, MNHN); same locality, viii.1984, [S.] Pokorný leg., 1 \(\text{(JRUC)}; \) \(\text{Meghalaya state} \): Khassia Hills [centroid ca. 25°30'N 091°30'E], 4000 ft [ca. 1220 m], 1892, Ex Musaeo H. W. Bates, 1 ♀ (coll. R. Oberthür, MNHN); Rajasthan state: Keoladeo National Park [ca. 27°10'N 077°30'E], 29.x.1997, J. Šťastný leg., semi-dry carrion of mongoose (Herpestes sp.), on dry soil trampled by cows, $4 \, \text{??}$, $1 \, \text{?}$ (JRUC); <u>Union Territory of Puducherry</u>: Pondichéry [= Pondicherry, ca. 11°56'N 079°50'E], without date and collector's name, 1 $\stackrel{?}{\circ}$ (coll. M. Pic, MNHN); Pondich.[erry], without date, Ex Musaeo Guér.-Menev. [= F.É. Guérin-Méneville], 3 👌 (coll. R. Oberthür, MNHN); Uttarakhand state [= Uttaranchal]: Kumaon, Haldwani division [ca. 29°13'N 079°31'E], without date, H. G. C.[hampion] leg., 1 ♀ (BMNH); Dehra Dun [ca. 30°20'N 078°02'E], vi.1944, without collector's name, $1 \circlearrowleft 1 \hookrightarrow (ZSM)$; same data, $1 \circlearrowleft (JRUC)$; Uttar Pradesh state: Moradabad [ca. 28°50'N 078°46'E], without date and collector's name, 1 & (coll. A. Grouvelle, MNHN); West Bengal state: "Bengale" [centroid ca. 23°00'N 087°40'E], without date [but before 1825], [A.] Duvaucel leg., 1 & (MNHN);

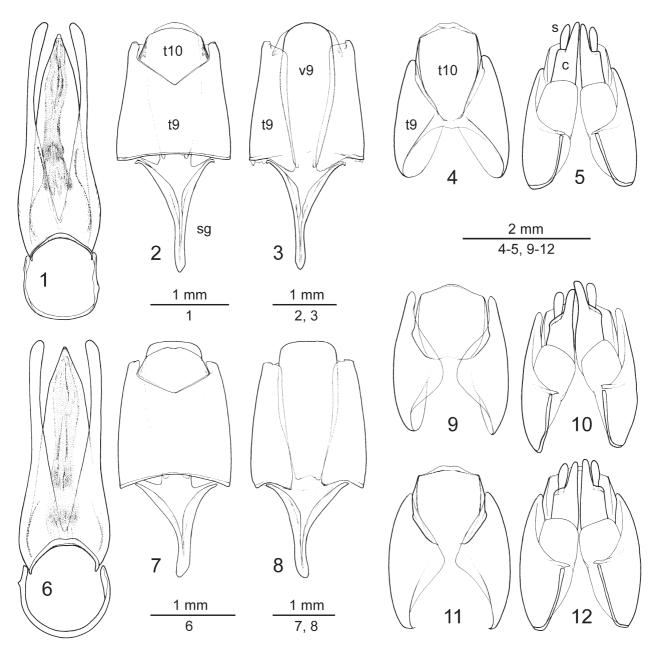
Nepal: Bara district, Amelkhganj [ca. 27°17'N 084°59'E], 520 m, viii.1957, E.I. Coher leg., $1 \subsetneq$ (FMNH); Chitwan district, Chitwan National Park [ca. 27°30'N 084°20'E], Saura [= Sauraha], 23.v.1988, S. Bílý leg., $1 \subsetneq$ (JRUC); "Nepaul", without date and collector's name, Collection Laporte, $1 \circlearrowleft$ (coll. M. Pic, MNHN); "Nepaul", without date [but before 1922] and collector's name, $1 \circlearrowleft$ (coll. L. Bedel, MNHN); "Nepal", without date and collector's name, $1 \circlearrowleft$ (OUMNH);

Diagnostic description. Body length 17.5–22.3 mm (21.0 mm in the lectotype of *Silpha rufithorax*), maximum body width 8.9–11.6 mm (11.1 mm in the lectotype of *S. rufithorax*). Body much wider than in *N. luciae* **sp. nov.**, widely rounded posteriorly in dorsal view (Figs. 15–16). Pronotum widely hexagonal in shape (Fig. 22), 1.63–1.78 times as wide as long. Ventrite 8 on abdomen black, with pale transverse line along posterior margin.

Male. Apex of elytron truncate, with posterior portion sinuous, suture extended in small, distinct inner denticle (Fig. 19). Genital segment in dorsal view with deeply notched suture between tergum 9 and 10, forming rectangular angle (Fig. 2); slender ventrite 9 with regularly rounded posterior margin (Fig. 3). Spiculum gastrale is slender, of equal length as tergum 9 (Figs. 2–3). Length of aedeagus 3.8 mm, median lobe more slender than in *N. luciae* **sp. nov.**; gently tapering to more elongated apical portion (Fig. 1). Parameres subapically more slender than in *N. luciae* **sp. nov.** (Fig. 1). Basal portion of aedeagus forming slender, sub-quadrate rim (Fig. 1).

Female. Elytron with elongate, convex apex, suture extended in small, but distinct inner denticle (Fig. 20). Tergum 10 elongate, tergum 9 reaching only two-thirds of tergum 10 length in dorsal view (Fig. 4).

Notes on taxonomy and nomenclature. *Oiceoptoma tetraspilotum* Hope, 1833 is confirmed here as conspecific junior subjective synonym of *Silpha rufithorax* Wiedemann, 1823. Two syntypes of *S. rufithorax* from ZMUC were designated here as lectotype (the male) and paralectotype (the female), in order to ensure the consistent application of the name, according to the Article 74.7.3 of the Code (ICZN 1999).

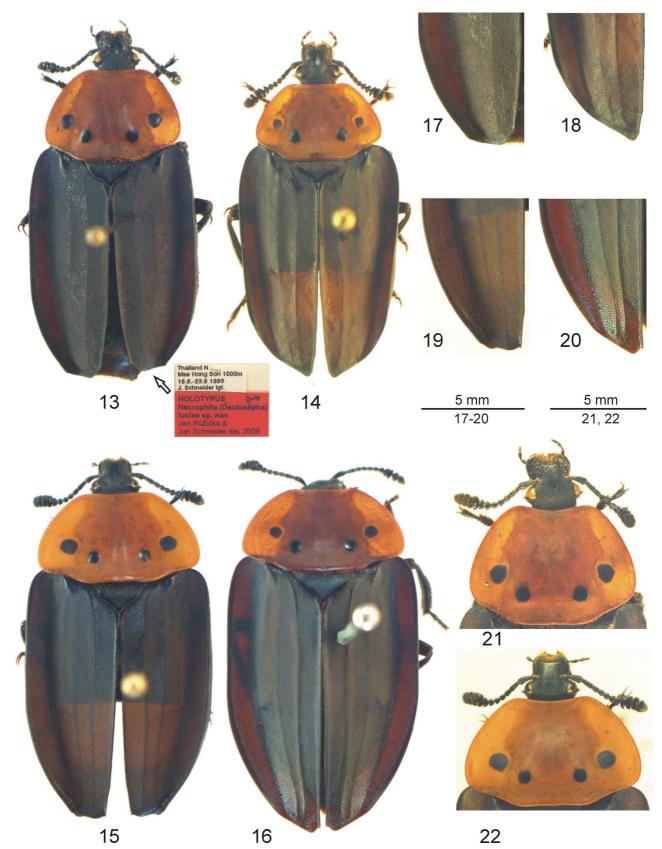


FIGURES 1–12. Genitalia of *Necrophila (Deutosilpha) rufithorax* (Wiedmann) (1–5, India: Keoladeo National Park, JRUC) and *N. (D.) luciae* Růžička & Schneider, **sp. nov.** (6–8, holotype ♂, BMNH, and 9–12, paratypes ♀♀, JRUC, from Thailand: Mae Hong Son), setae omitted. Aedeagus ventrally (1, 6); ♂ genital segments dorsally (2, 7) and ventrally (3, 8); ♀ genital segments dorsally (4, 9, 11) and ventrally (5, 10, 12). Abbreviations: c—coxite, s—stylus, sg—spiculum gastrale, t9—tergum 9, t10—tergum 10, v9—ventrite 9.

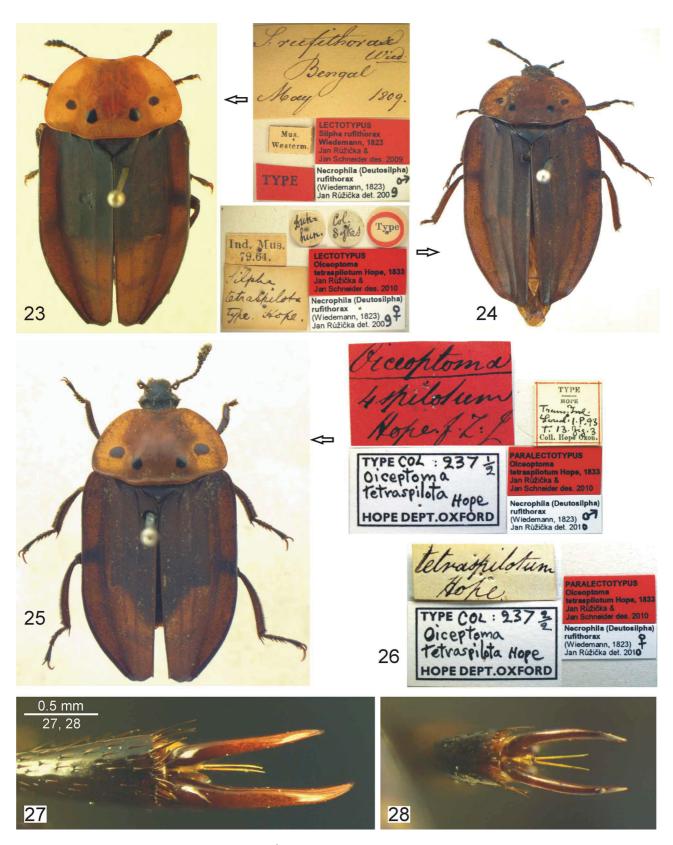
The syntype of *Oiceoptoma tetraspilotum* deposited in BMNH is designated here as a lectotype, in order to ensure the consistent application of the name, according to the Article 74.7.3 of the Code (ICZN 1999).

Additional two syntypes (a male and a female) from OUMNH are designated as paralectotypes. Hope (1934: 93) mentioned "circa Poona copiosè [= abundant] / Mus. Sykes", not providing exact number of specimens examined. The lectotype specimen from BMNH bears the labels "Col. Sykes" and "Ind. Mus. / 79.64" (Fig. 24), which indicate that it was part of the W.H. Sykes collection in India and become part of the BMNH collection only in 1879. The additional two specimens from OUMNH were probably retained by Hope in his collection.

The nomenclature of *Oiceoptoma tetraspilotum* is interesting: the species was described twice. Preliminar (but valid) diagnosis was published by Hope (1833) in *Proceedings of the Zoological Society of London*, forming part of [W.H.] Sykes' report of the Society meeting. Full description was published by Hope (1834) in *Transactions of the Zoological Society of London*, together with a colour habitus figure and details of the head and antenna. As the first



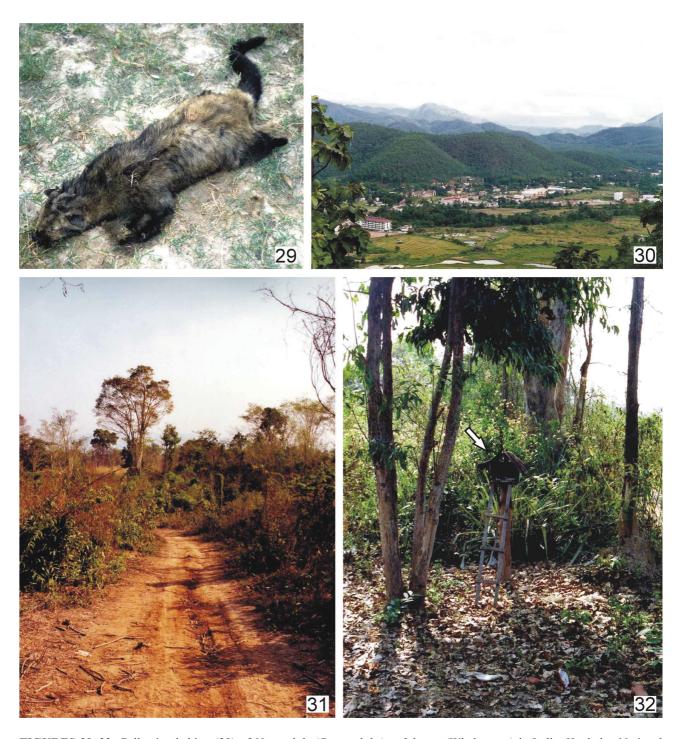
FIGURES 13–22. Habitus dorsally of *Necrophila (Deutosilpha) luciae* Růžička & Schneider, **sp. nov.** (13, 17, 21, holotype \circlearrowleft , BMNH, and 14, 18, paratype \Lsh from Thailand: Mae Hong Son, JRUC; body length 22.1 mm in \circlearrowleft and 22.7 mm in \Lsh) and *N.* (*D.*) *rufithorax* (Wiedmann) (15, 19, 22, \circlearrowleft from India: Keoladeo National Park, JRUC; and 16, 20, \backsim from India: Poona, BMNH; body length 20.6 mm in \circlearrowleft and 20.8 mm in \backsim). Habitus dorsally (13–16), apex of left elytron dorsally (17–20), \circlearrowleft pronotum in orthogonal view (21–22).



FIGURES 23–28. Habitus dorsally and labels of \circlearrowleft lectotype of *Silpha rufithorax* Wiedemann (23, "circa Poona", ZMUC), \updownarrow lectotype of *Oiceoptoma tetraspilotum* Hope (24, "Dukhun", BMNH) and \circlearrowleft paralectotype of *O. tetraspilotum* (25, OUMNH), and labels of \updownarrow paralectotype of *O. tetraspilotum* (26, OUMNH). Metatarsus in dorsal view of *Necrophila (Deutosilpha) rufithorax* (Wiedemann) (India, Keoladeo National Park, JRUC): male (27) and female (28).

paper is in fact only a report, which summarize the diagnoses (of several species of Coleoptera from Deccan plateau) and mentioned colour plates presented during the meeting and published a year later, we consider unnecessary treating both names of 1833 and 1834 as separate homonyms.

Collecting circumstances. Series of specimens from India: Keoladeo National Park was collected on semi-dry carcass of mongoose (*Herpestes* sp., Herpestidae) (Fig. 29), on dry soil trampled by cows, in the middle of field (J. Šťastný, pers. comm.).



FIGURES 29–32. Collecting habitat (29) of *Necrophila (Deutosilpha) rufithorax* (Wiedemann) in India: Keoladeo National Park, on carrion of mongoose (*Herpestes* sp.); landscape type (30) of *N. (D.) luciae* Růžička & Schneider, **sp. nov.** in Thailand: Mae Hong Son; landscape type (31) and collecting habitat (32) of *N. (D.) luciae* Růžička & Schneider, **sp. nov.** in Laos: Ban Thabok, construction with pitfall trap on its top is indicated by arrow (photo 29 by J. Šťastný, 30 by J. Schneider, 31–32 by J. Kantoh).

Distribution. Widely distributed through the Indian Peninsula, south of the main Himalayan mountain ridge. So far known from India, Nepal and Sri Lanka (the last one without more precise locality) (Fig. 33). Published records from Thailand (Růžička *et al.* 2000) and Laos (Růžička & Schneider 2003) concern the following species.

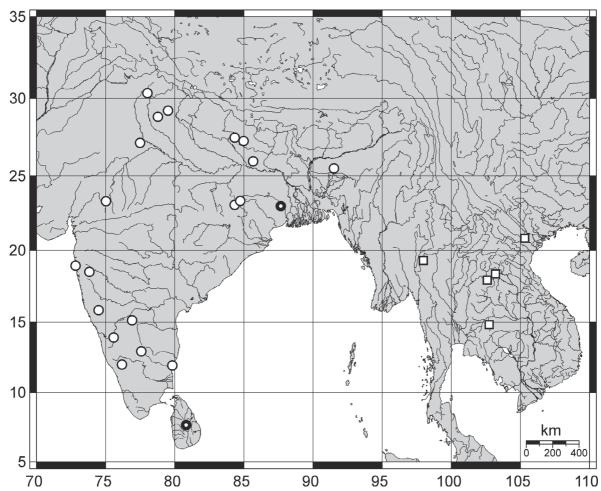


FIGURE 33. Known distribution of *Necrophila (Deutosilpha) rufithorax* (Wiedmann) (open circles; thick margins for state records only) and *N. (D.) luciae* Růžička & Schneider, **sp. nov.** (open squares; Chinese record from "Ouy-Sy" not figured) in Oriental region.

Necrophila (Deutosilpha) luciae Růžička & Schneider, sp. nov. (Figs. 6–14, 17–18, 21, 33)

Deutosilpha rufithorax: Růžička et al. 2000: 382 (record from Thailand, misidentification) Deutosilpha rufithorax: Růžička & Schneider 2003: 309 (record from Laos, misidentification)

Type locality. Thailand, Mae Hong Son, 1000 m.

Type material. 67 specimens—Holotype \circlearrowleft (BMNH), labelled (Fig. 13): "Thailand N / Mae Hong Son [ca. 19°18'N 097°58'E] 1000m / 16.6.–23.6. 1993 / J. Schneider lgt. [p] // HOLOTYPUS [p] \circlearrowleft [hw] / Necrophila (Deutosilpha) / luciae **sp. nov.** / Jan Růžička & / Jan Schneider det. 2009 [p, red label]". Paratypes: 2 \circlearrowleft \circlearrowleft 2 \circlearrowleft (FMNH): same data as holotype, except for "PARATYPE [p]" and " \circlearrowleft [hw]" or " \supsetneq [hw]"; 4 \circlearrowleft \circlearrowleft 6 \circlearrowleft (JHAC): same data; 2 \circlearrowleft 2 \circlearrowleft (JMIC): same data; 4 \circlearrowleft 4 \circlearrowleft (JRUC): same data; 12 \circlearrowleft (SMNS): same data; 1 \circlearrowleft (NMPC): same data; 1 \circlearrowleft (SMLC): same data; 1 \circlearrowleft (Paratypes) (MTDC); same data; 1 \circlearrowleft (SMNS): same data; 2 \circlearrowleft \circlearrowleft (NMPC): same data; 1 \circlearrowleft (SMNS), labelled: "Thailand 25.VIII.1996 / Pak Thong/Khao Yai / 14°50'N 102°44'E / leg. M. HAUSER [p] // PARATYPUS [p] \supsetneq [hw] / Necrophila (Deutosilpha) / luciae **sp. nov.** / Jan Růžička & / Jan Schneider det. 2009 [p, red label]"; 1 \supsetneq (coll. R. Oberthür, MNHN), labelled: "Sutchuen [= China: Sichuan province] / Ouy-Sy, / R. P. Mombelg [leg.] / 1911 [p] // MUSÉUM PARIS / 1952 / COLL. R. OBERTHUR [p] // PARATYPUS [p] \supsetneq [hw] / Necrophila (Deutosilpha) / luciae **sp. nov.** / Jan Schneider det.

2009 [p, red label]"; 1 & (NHMB), labelled: "CHINA [p, label with a black frame] // Sammlung / G. Schneider / Museumsverein [p] // Necrophila (Deutosilpha) / luciae sp. nov. / Jan Růžička & / Jan Schneider det. 2009 [p, red label]"; 1 & (MNHN), labelled: "MUSEUM PARIS / TONKIN / RÉG: DE HOA BINH [= Vietnam, Hòa Bình, ca. 20°49'N 105°19'E] / A. DE COOMAN 1928 [p] // PARATYPUS [p] & [hw] / Necrophila (Deutosilpha) / luciae sp. nov. / Jan Růžička & / Jan Schneider det. 2009 [p, red label]"; 1 & 1 \(\frac{1}{2} \), 1 \(\text{Q (HNHM)} \), labelled "LAOS, Vientiane, / shore of Mekong River [ca. 17°57'N 102°36'E], / singled from faeces, // No. 4 / 12.–20.III. 1998 / leg. O. Merkl & G. Csorba [p] // Deutosilpha / rufithorax W. [hw] / det. Schawaller 1999 [p] // PARATYPUS [p] & [hw] [or \(\pi \)] / Necrophila (Deutosilpha) / luciae sp. nov. / Jan Růžička & / Jan Schneider det. 2009 [p, red label]"; 1 & 1 \(\frac{1}{2} \), 1 \(\pi \) (MNIC), labelled: "Ban Tabok [= Laos, Ban Thabok, ca. 18°22'N 103°12'E, ca. 154 m in alt.], Boli Khamxai Pref., 17–III–2003, J. Kantoh leg. [p] // PARATYPUS [p] \(\frac{1}{2} \) [hw] [or \(\pi \)] / Necrophila (Deutosilpha) / luciae sp. nov. / Jan Růžička & / Jan Schneider det. 2009 [p, red label]"; 1 \(\pi \) (TFUC), same locality and date, but "Raiichiro Katsuyama leg. // PARATYPUS [p] \(\pi \) [hw] / Necrophila (Deutosilpha) / luciae sp. nov. / Jan Růžička & / Jan Schneider det. 2009 [p, red label]"; 1 \(\frac{1}{2} \) (aedeagus mounted on a small card) (coll. M. Pic, MNHN), labelled: "Muséum Paris / Coll. Pic [p, yellow label] // genre / Deutosilpha Port. [hw, Pic's ms, orange label] // PARATYPUS [p] \(\frac{1}{2} \) [hw] / Necrophila (Deutosilpha) / luciae sp. nov. / Jan Růžička & / Jan Schneider det. 2009 [p, red label]".

Diagnostic description. Body length 18.9–22.5 mm (22.1 mm in the holotype), maximum body width 8.9–10.6 mm (10.4 mm in the holotype). Body more slender than in *N. rufithorax*, elytra subquadrate in dorsal view (Figs. 13–14). Pronotum hexagonal in shape (Fig. 21), 1.35–1.57 times as wide as long. Ventrite 8 on abdomen black, with pale triangular macula medially in posterior third.

Male. Apex of elytron subtruncate, with regularly rounded posterior portion (Fig. 17). Genital segment in dorsal view with less deeply notched suture between tergum 9 and 10 than in *N. rufithorax*, forming obtuse angle (Fig. 7); ventrite 9 more stout than in *N. rufithorax*, with subquadrate posterior margin (Fig. 8). Spiculum gastrale is more stout than in *N. rufithorax*, distinctly shorter than the length of tergum 9 (Figs. 7–8). Length of aedeagus 3.5 mm, median lobe wider than in *N. rufithorax*, stout; with short, triangular apical portion (Fig. 6). Parameres more stout than in *N. rufithorax* (Fig. 6). Basal portion of aedeagus forming robust, regularly rounded rim (Fig. 6).

Female. Elytron with triangular, flat apex, suture extended in only with very vestigial inner denticle or without denticle (Fig. 18). Tergum 10 oval, tergum 9 reaching posterior margin of tergum 10 (Fig. 9); exceptionally, reaching only three-quarters of tergum 10 length (Fig. 11).

Collecting circumstances. The large series from the type locality was found on the periphery of the town, in a garden with sparse vegetation close to the slaughterhouse, individually collected on numerous decaying cow heads; the surrounding hills are covered by a dense forests (Fig. 30). The two specimens from Laos: Vientiane were collected from excrements, on the shore of Mekong river. The three specimens from Laos: Ban Thabok were caught by a carrion trap using the dead body of a small mammal resembling a squirrel as the bait; the baited trap was covered by a roof like dwarf' house at ca. 1.5 m above the ground (Fig. 32) inside the sparse grove along dry rice fields situated between a river and a private house; the trap site was relatively dry including its surroundings (Fig. 31). The beetles were found together with saprinine histerids, scarabaeids, trogids and *Diamesus osculans* (Vigors, 1825) (Silphidae) two days after the trap was set (J. Kantoh, pers. comm. to M. Nishikawa). It is no doubt that all three specimens from Laos: Ban Thabok perform active flight.

Etymology. Patronymic, named after Lucie Schneiderová, the older daughter of the second author.

Distribution. So far known from Thailand, Laos, Vietnam and China: Sichuan province (the last one with only unprecisely located, single record) (Fig. 33).

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